

Original Research Article

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Detection of Multidrug Resistance and Characterization of Mutations in *Mycobacterium tuberculosis* Isolates in Raichur District, India

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ABSTRACT

Tuberculosis remains as one of the leading cause of morbidity and mortality globally. Estimated cases of around 9.6 million are affected by all forms of TB worldwide, out of which 2.2 billion are affected in India. India also has the second highest burden of MDR TB after China. Information on MDR TB is sparse in Raichur District. We undertook this study to detect MDR TB among MDR suspected cases and its common mutation using Genotype MTBDR plus. All presumptive MDR sputum samples were sent to DST LAB RIMS, Raichur from various part of Raichur district. Samples in study were taking during the period from January 2017 to April 2017. Line Probe Assay was done on sputum samples to detect common mutations in *rpoB* gene for Rifampicin and *katG* gene and *inhA* gene for Isoniazid resistance respectively. Out of the 448 samples received from MDR suspects, 49(10.9%) were found to be MDR TB. Commonest pattern for Rifampicin resistance was missing WT8 probe along with mutation in MUT3 band (S531L) codon, and mutation of MUT1 band (S5315T) of *katG* gene was commonest pattern for Isoniazid resistance. The MDR-TB among presumptive MDR in Raichur district was found to be 10.9%, and the mutation pattern obtained for Rifampicin and Isoniazid resistance were similar to those reported earlier.

Keywords

MDR-TB, Drug resistance, Line probe assay, Rifampicin and Isoniazid, Drug susceptibility testing.

Article Info

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Introduction

TB in India is an ancient disease, and in Indian literature it has been mentioned that it occurred from around 1500 BCE. But at the beginning of the nineteenth century it was generally thought that there was hardly any TB occurring in India, so it was the least area of concern, It was in The second All-India Sanitary Conference, Madras 1912, due to increase mortality caused by TB there was discussion of TB and different views were expressed to the measures that should be taken. Like formation of anti-tuberculosis societies, the establishment of more sanatoria,

the establishment of dispensaries, the improved ventilation of homes and schools, as well as the use of tuberculin¹. Scenario has changed now in regard to TB.

According to WHO report in 2015, there were an estimated 10.4 million new (incident) TB cases worldwide Six countries accounted for 60% of the new cases: India, Indonesia, China, Nigeria, Pakistan and South Africa.²

In MDR TB are resistance to Rifampicin, with or without resistance to other drugs²,

XDR is defined as resistance to Rifampicin and Isoniazid as well as any member of Quinolone and at least one of the second line anti TB injectable drugs.

In 2015, there were an estimated 4,80,000 new cases of multidrug-resistant TB (MDR-TB).²

India, China and the Russia accounted for 45% of the combined total of 5,80,000 cases. India has second highest TB burden MDR-TB rates have been found to be 17.4 to 53 per cent among previously treated cases who are more likely to develop multi-drug resistance. Reportedly first case of XDRTB is also detected in India in November 2011. (3, 4, 5)

This kind of rapid progression of drug resistance from MDR to XDR and XXDR TB underline the needs for rapid and accurate diagnosis of drug resistance TB. Newer diagnostic tests like Cartridge Based Nucleic Acid Amplification Test (CB-NAAT), MGIT 960 and Line Probe Assay (LPA) are some of the tests for the diagnosis of tuberculosis rapid detection of RIF and INH resistance. XDR was first reported in 2006, since then it has spread to 6 continent and 55 countries (2). This kind of rapid progression of drug resistance from MDR to XDR and XXDR TB underline the needs for rapid and accurate diagnosis of drug resistance TB.

The diagnosis of MDR TB is based on isolation of strain resistance to medicines. Newer diagnostic tests like Cartridge Based Nucleic Acid Amplification Test (CB-NAAT), MGIT 960 and Line Probe Assay (LPA) are some of the tests for the diagnosis of tuberculosis and rapid detection of RIF and INH resistance.

The data for MDR TB is sparse in this region of the state due to lack of proper infrastructure and training, Hence we have undertaken this study for detection of MDR

TB and common mutation among MDR suspects using Line Probe Assay, this study could be useful for strategic planning and control of MDR TB in this region of state.

Materials and Methods

All presumptive MDR patients' sputum samples were collected during the period from January 2017 to April 2017 to DST LAB Department of Microbiology, RIMS Raichur.

MDR suspect criteria includes

Failure

Re-treatment cases sputum positive at 4th months.

Sputum positive cases at diagnosis, re-treatment cases.

Any follow up sputum positive

Sputum negative at diagnosis, re-treatment case

HIV TB cases Line Probe Assay was done on these suspected samples

Sample preparation and processing (NaCl-NaOH method)

All the sample preparation and processing was done in BSL 3 laboratory. The samples were processed using the working solution of 50ml of 4% Sodium hydroxide (NaOH)+50 ml of 2.9% Sodium citrate + 500 mg of NALC powder.

DNA extraction was done using Lysis A and Neutralizing Buffer, One part used for amplification and rest was stored at -20 degree C. Master mix was prepared using the reagent provided in the kit 5microlitre DNA was added to 45 microlitre of master mix

solution Then DNA Amplification and Hybridization was carried out per manufactures instruction DNA extraction, Master mix preparation, DNA amplification and Hybridization all were carried out in separate designated rooms and unidirectional workflow was maintained. The results of DNA strips were interpreted with the help of reporting card (Fig. 2). The absence or presence of wild type and mutant band were recorded.

Results and Discussion

Out of 448 samples in which Line Probe Assay was performed (Fig. 1), Resistance to both Rifampicin and Isoniazid was found in 33/448 (7.36%) isolates, mono resistance to Rifampicin was found in 16/448 (3.6%) isolates, and mono resistance to Isoniazid was found in 31 (6.9%) isolates, Total Rifampicin resistance was found to be 49/448 (10.9%). Remaining 368 (82.2%) isolates out of 448 were found to be susceptible to both Rifampicin and Isoniazid.

Among 49 total Rifampicin resistance isolates, missing wild type (WT) probe with known mutation was found in 29/49 (59.1%) isolates, *rpoB* WT8(531-533) was the commonest wild type probe missing (25/29,86.2%) followed by WT7(3/29,10.3%) and WT1(1/29) (Table 1).

Commonest Rifampicin mutation in these strains was in S531L codon (25/29, 86.2%) followed by mutation in H526Y codon (2/29, 6.8%), then in H526D and D516V codon both having 1/29(3.4%) mutation (Table 2). In 6/49(12.4%) Rifampicin resistance strains, one or more wild type (WT) probes were missing with no mutation band found in them. In 5/49(10.3%) Rifampicin resistance strain, one or more wild type probe was missing with mutation band found on them. WT3/WT4 were the most common wild type band missing in these type was strain (5/49).Mixed

pattern of Rifampicin resistance in which all wild type (WT) of probe were present along with the presence of one or more mutant band was present in 8/49(16.3%) isolates (Table 1), MUT3(S531L) was the most common mutation present (7/8,87.5%) followed by MUT1(1/8,12.5%) (Table 2).

Out of 448 samples total of 64(14.2%) Isoniazid resistant isolates were detected. Missing wild type *katG* WT1 was most common probe missing with known mutation 42/64 (65.6%) (Table 3), most common mutation in *katG* was found in MUT1 band (S315T1 codon) in 42/42(100%) strains.

In 2/64(3.1%) strains of Isoniazid resistance wild type *katG*WT1 probe was missing with no mutation band (Table 3), mixed *katG* resistance was found in 6/64(9.3%) samples with MUT1 band (S315T1 codon) mutation in 6/6 (100%) strains (Table 4) *inhA* gene with missing wild type probe *inhA*WT1 with known mutation was found in 9/64(14.5%) (Table 3), 9/9(100%) *inhA* MUT1 band C15T codon was most common mutation present in these strain. with missing wild type which included 14/14 (100%) mutation *inhA* MUT1 in C15T codon. Both *inhA* and *katG* mutation was found in 2/64(3.1%). Mixed pattern of Isoniazid resistance was found in 1/64(1.6%) isolates (Table 4).

Out of total 448 isolates 49 (10.9%) were found to be MDR TB, Commonest pattern for Rifampicin resistance was missing WT8 probe with mutation in codon S531L being most common and missing of *katG* gene was commonest pattern for Isoniazid resistance with mutation in S53T1 was most common.

Similar study was done by Ritu Singhal *et al.*, in Northeast state of India during the period of January 2012 to august 2012, in which from total of 553 sputum samples, 181 (32.7%) isolates were found to be multidrug resistant. Missing WT8 along with mutation

in codon S531L was commonest pattern for Rifampicin resistant isolates (65.1%) and missing WT along with mutations in codon

S315T1 of *katG* gene was commonest pattern for Isoniazid resistant isolates (86.2%).⁽⁶⁾

Fig.1 Methodology of line probe assay

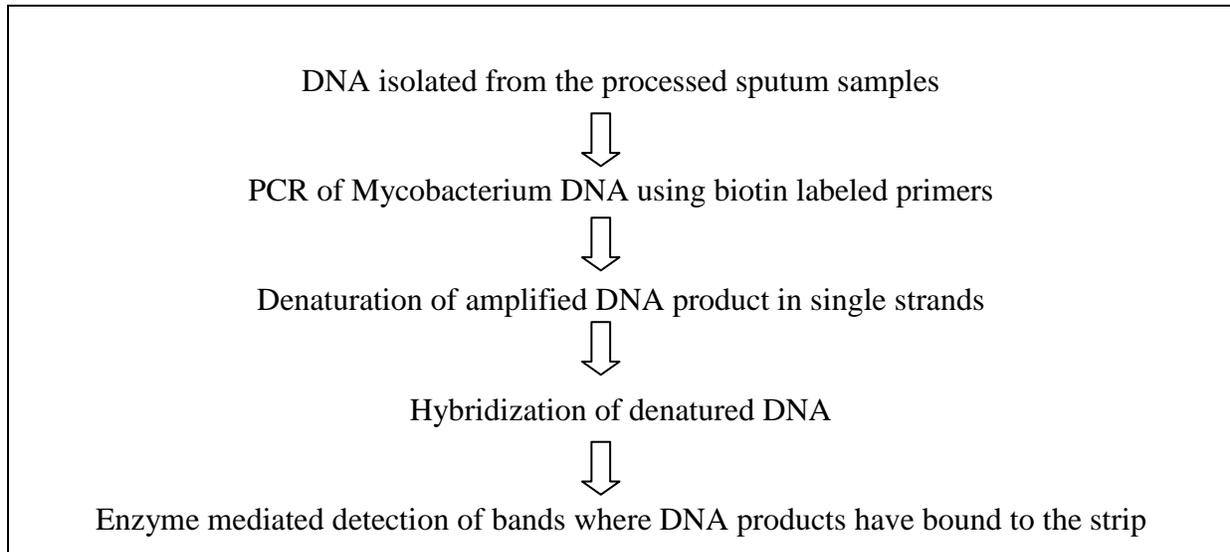


Fig.2 Reporting card for line probe assay

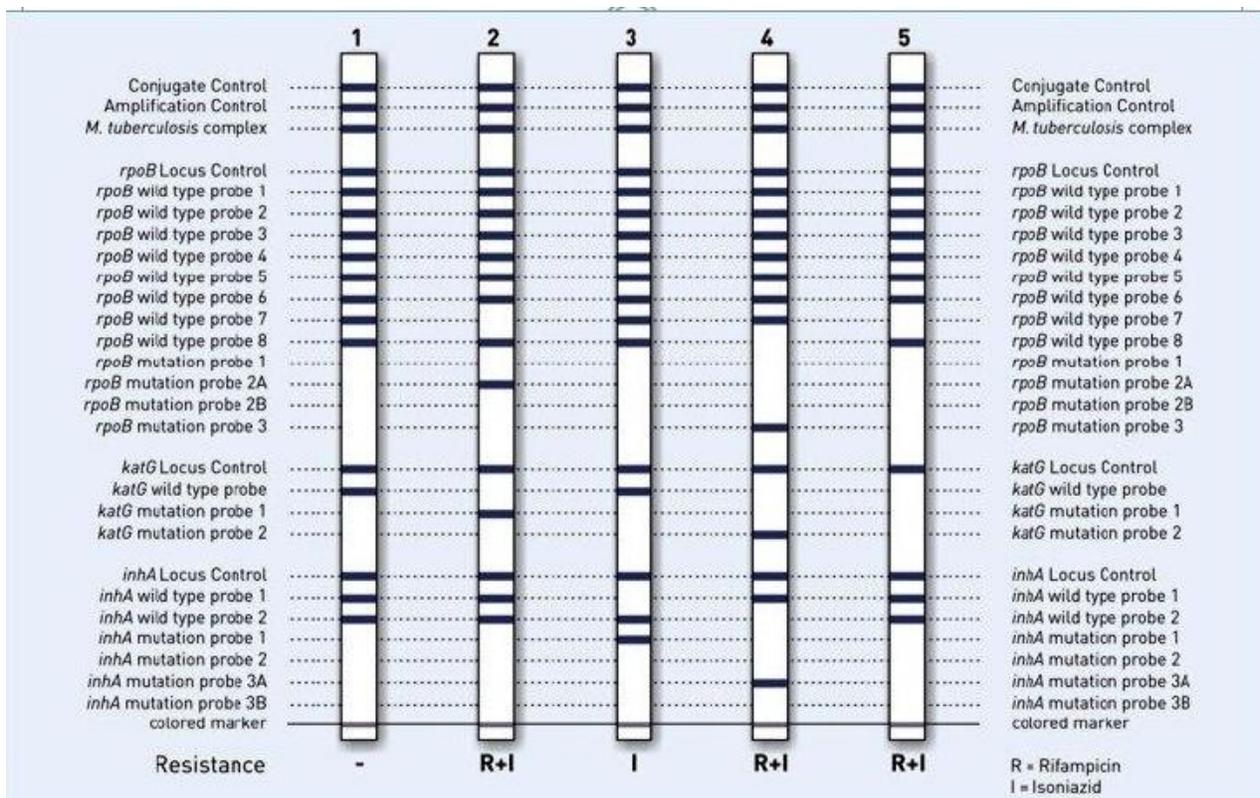


Table.1 Missing wild type of probe for Rifampicin resistance.
Total Rifampicin resistance isolates (n-49)

GENE	BAND MISSING	MISSING WILD TYPE BAND WITH KNOWN MUTATION n-29	ONE OR MORE MISSING WILD TYPE BAND WITH NO MUTANT PROBE n-6	ONE OR MORE MISSING WILD TYPE BAND WITH MUTANT PROBE n-5
<i>rpoB</i>	WT1	01	00	00
	WT2	00	02	00
	WT3	00	02	05
	WT4	00	01	05
	WT5	00	01	00
	WT6	00	00	00
	WT7	03	02	00
	WT8	25	02	00

Table.2 Mutation pattern in Rifampicin resistance. Total Rifampicin resistance isolates (n-49)

MUTATION PRESENT	IN MISSING WILD TYPE WITH KNOWN MUTATION n-29	IN ONE OR MORE WILD TYPE MISSING WITH MUTANT PROBE PRESENT n-5	IN ONE OR MORE WILD TYPE MISSING WITH MUTANT PROBE ABSENT n-6	MIXED RIFAMPICIN RESISTANCE n-8
<i>rpoBMUT1 (D516V)</i>	01	04	00	01
<i>rpoBMUT2A (H526Y)</i>	02	00	00	00
<i>rpoBMUT2B(H526D)</i>	01	00	00	00
<i>rpoMUT3(S531L)</i>	25	01	00	07

Table.3 Missing wild type of probe for Isoniazid resistance. Total Isoniazid isolates (n-64)

GENE	BAND MISSING	WILD TYPE MISSING WITH KNOWN MUTATION	WILD TYPE MISSING WITH NO MUTANT BAND
<i>KatG</i>	WT	42	02
<i>inhA</i>	WT1	09	02
	WT2	00	00

Table.4 Mutation pattern in Isoniazid resistance. Total Isoniazid resistance isolates (n-64)

MUTATION PRESENT	IN MISSING WILD TYPE WITH KNOWN MUTATION	IN ONE OR MORE WILD TYPE BAND MISSING WITH MUTANT PRESENT	IN ONE OR MORE WILD TYPE MISSING WITH NO MUTANT PROBE n-4	MIXED ISONIAZID RESISTANCE	MUTATION PRESENT IN BOTH <i>inhA</i> AND <i>katG</i> GENE n-1
<i>katGMUT1 (S315T1)</i>	42	00	00	06	01
<i>katGMUT 2 (S315T2)</i>	00	00	00	00	00
<i>inhAMUT1 (C15T)</i>	09	00	00	02	01
<i>inhAMUT 2 (A16G)</i>	00	00	00	00	00
<i>inhAMUT 3A (T8C)</i>	00	00	00	00	00
<i>inhA MUT 3B (T8A)</i>	00	00	00	00	00

A comparison study between Xpert MTB/RIF with Line Probe Assay for rapid detection of Rifampicin Mono-resistant Mycobacterium tuberculosis was done by Syed Beenish Rufai and *et al.*, in October 2013. In a total of 405 sputum samples, 285 smear positive samples were subjected to LPA. 72(25.8%) samples showed multidrug resistance, 62(22.2%) showed Rifampicin mono-resistance, 29 (10.3%) showed Isoniazid mono-resistance, Six (2.1%) of the samples gave invalid results.

Of the 62 Rifampicin Mono-resistant samples by LPA, 38 (61.4%) showed Rifampicin resistance, while 21 (33.8%) were found susceptible to Rifampicin by Xpert MTB. The MGIT960 results showed 100% agreement with LPA results but only 64.4% agreement with Xpert MTB/RIF results. Sequencing analysis of discrepant samples showed 91.3% concordance with LPA but only 8.7% concordance with the Xpert MTB/RIF assay⁽¹⁰⁾. These findings clearly indicate that more advancement has to be done in the case for Xpert MTB so that its sensitivity can be increased for rapid diagnosis.

Our area of concern is that Cases of pulmonary tuberculosis have been missed along with the case of MDR TB, and progression of disease from TB which can be cured in a period of 6-9 months to MDR TB which take upto 24 months to cure and the treatment course which also has many side effects is alarming. The reason of this progression is partly due to lack of infrastructure and failure to implement the policies.

According to National Strategic Plan 2012-2017 for TB, it has been decided to extend the reach of RNTCP services to peripheral regions to diagnose TB including MDR TB and expanded its services for management of multidrug resistance TB. These targets can be

achieved by rapid diagnosis of TB and drug susceptibility testing.²

Conventional drug susceptibility testing using solid media such as LJ media is time consuming, it takes upto 3 months. Liquid culture medium MGIT 960 are sensitive and faster (average time 14 days)⁷ for detection of MDR TB whereas Line Probe Assay have a rapid turnover time of 48-72 hours for detection of MDR TB.⁸

Line Probe Assay has also been approved by WHO⁸ for use in low income setting, so it can be used as rapid detection of MDR TB and common mutation for the resistance can be detected⁹ and early treatment can be started.

After setting up of DST lab in Raichur Institute of Medical Sciences, Raichur Microbiology Department, Line Probe Assay could be very helpful for the detection of MDR TB in this North Karnataka region and will help to decrease the burden of TB on the society.

References

1. WHO | Tuberculosis [Internet]. WHO. [cited 2016 Sep 23]. Available from: <http://www.who.int/mediacentre/factsheets/fs104/en/>
2. TB Facts | TB, tests, drugs, statistics [Internet]. TB Facts.org.[cited 2016 Sep 23]. Available from: <http://tbfacts.org/>
3. Ramachandran R, Nalini S, Chandrasekar V, Dave PV, Sanghvi AS, Wares F, et al. Surveillance of drug-resistant tuberculosis in the state of Gujarat, India. *Int J Tuberc Lung Dis*. 2009 Sep;13(9):1154–60.
4. Hanif M, Malik S, Dhingra VK. Acquired drug resistance pattern in tuberculosis cases at the State

- Tuberculosis Centre, Delhi, India. *Int J Tuberc Lung Dis.* 2009 Jan;13(1):74–8.
5. Paramasivan CN, Rehman F, Wares F, Sundar Mohan N, Sundar S, Devi S, et al. First- and second-line drug resistance patterns among previously treated tuberculosis patients in India. *Int J Tuberc Lung Dis.* 2010 Feb;14(2):243–6.
 6. Singhal R, Myneedu VP, Arora J, Singh N, Sah GC, Sarin R. Detection of multi-drug resistance & characterization of mutations in *Mycobacterium tuberculosis* isolates from North- Eastern States of India using GenoType MTBDRplus assay. *Indian J Med Res.* 2014 Oct;140(4):501–6.
 7. Somoskövi A, Ködmön C, Lantos A, Bártfai Z, Tamási L, Füzy J, et al. Comparison of recoveries of mycobacterium tuberculosis using the automated BACTEC MGIT 960 system, the BACTEC 460 TB system, and Löwenstein-Jensen medium. *J Clin Microbiol.* 2000 Jun;38(6):2395–7.
 8. World Health Organisation: Policy Statement. Molecular Line Probe Assays for Rapid Screening of patients at risk of multidrug resistant tuberculosis (MDR-TB), 2008. Accessed 18 November 2000.
 9. Brossier F, Veziris N, Truffot-Pernot C, Jarlier V, Sougakoff W. Performance of the genotype MTBDR line probe assay for detection of resistance to rifampin and isoniazid in strains of *Mycobacterium tuberculosis* with low- and high-level resistance. *J Clin Microbiol.* 2006 Oct; 44(10):3659–64.
 10. Syed Beenish Rufai, Parveen Kumar, Amit Singh, Suneel Prajapati, Veena Balooni, and Sarman Singh. Comparison of Xpert MTB/RIF with Line Probe Assay for Detection of Rifampin-Monoresistant *Mycobacterium tuberculosis*. *J Clin Microbiol.* 2014 Jun; 52(6): 1846–1852.

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